AMENDMENT TO THE CLAIMS

This listing of claims will replace all prior versions of claims in the application.

Listing of Claims:

1. (Original) A method for increasing or maintaining the number of functional cells of a predetermined type in an organ or tissue of a mammal, wherein said organ or tissue is injured, damaged, or deficient in said functional cells, said method comprising administering to said mammal a composition enriched in pluripotent cells that express the Hox11 gene.

2. (Original) The method of claim 1, further comprising stimulating said organ or tissue before administering said composition.

3. (Original) The method of claim 2, wherein said organ or tissue is stimulated by administering TNF-alpha.

4. (Original) The method of claim 2, wherein said organ or tissue is stimulated by administering a TNF-alpha agonist or a TNF-alpha inducing substance.

5. (Previously Presented) The method of claim 4, wherein said TNF-alpha agonist or TNF-alpha inducing substance is Complete Freund's Adjuvant (CFA), ISS-ODN, microbial cell wall components with LPS-like activity, cholera particles, *E. coli* heat labile enterotoxin, *E. coli* heat labile enterotoxin complexed with lecithin vesicles, ISCOMS-immune stimulating complexes, polyethylene glycol, poly(N-2-(hydroxypropyl)methacrylamide), synthetic oligonucleotides containing CpG or CpA motifs, monophosphoryl lipid A, Bacillus Clamette-Guerin, γ-interferon, Tissue Plasminogen Activator, LPS, Interleukin-1, Interleukin-2, UV light, a lymphotoxin, cachectin, a TNFR-2 agonist, an intracellular mediator of the TNF-alpha signaling pathway, a NFκB inducing substance, IRF-1, STAT1, a lymphokine, a tumor necrosis factor-alpha (TNF-α) receptor II agonist, or the combination of TNF-alpha and an anti-TNFR-1 antibody.

6. (Original) The method of claim 5, wherein said TNF-alpha agonist or TNF-alpha inducing substance is Complete Freund's Adjuvant, Bacillus Clamette-Guerin, or γ-interferon.

7. (Original) The method of claim 2, wherein said organ or tissue is stimulated 6-12 hours before administering said composition.

8. (Original) The method of claim 1, wherein said composition is enriched in cells which do not express CD45 protein.

9. (Original) The method of claim 8, wherein said pluripotent cells are enriched from the peripheral blood or tissue of a mammal by a method comprising: a) providing from the mammal peripheral blood or tissue that contains pluripotent cells; b) separating pluripotent cells from said peripheral blood or tissue; c) separating said pluripotent cells into a first cell population which expresses CD45 antigen on the surface of said cells and a second cell population which predominantly does not express CD45 antigen on the surface of said cells; and d) selecting said second cell population.

10. (Original) The method of claim 1, wherein said pluripotent cells are derived from the spleen.

11. (Original) The method of claim 1, wherein said pluripotent cells are semi-allogeneic.

12. (Original) The method of claim 1, wherein said pluripotent cells are isogeneic.

13. (Withdrawn) A method for increasing or maintaining the number of functional cells of a predetermined type in an organ or tissue of a mammal, wherein said organ or tissue is injured, damaged, or deficient in said functional cells, said method comprising administering to said mammal a composition comprising pluripotent cells resulting from transfecting a pluripotent

or totipotent cell with a Hox11 gene.

14. (Withdrawn) The method of claim 13, wherein said Hox11 gene is expressed.

15. (Withdrawn) The method of claim 13, wherein said pluripotent cells are the result of transfecting a pluripotent cell.

16. (Withdrawn) The method of claim 15, wherein said pluripotent cell is semi-allogeneic.

17. (Withdrawn) The method of claim 15, wherein said pluripotent cell is isogeneic.

18. (Withdrawn) The method of claim 13, further comprising stimulating said organ or tissue before administering said composition.

19. (Withdrawn) The method of claim 14, wherein said organ or tissue is stimulated by administering TNF-alpha.

20. (Withdrawn) The method of claim 14, wherein said organ or tissue stimulated by administering a TNF-alpha agonist or a TNF-alpha inducing substance.

21. (Withdrawn) The method of claim 20, wherein said TNF-alpha agonist or TNF-alpha inducing substance is Complete Freund's Adjuvant (CFA), ISS-ODN, microbial cell wall components with LPS-like activity, cholera particles, *E. coli* heat labile enterotoxin, *E. coli* heat labile enterotoxin complexed with lecithin vesicles, ISCOMS-immune stimulating complexes, polyethylene glycol, poly(N-2-(hydroxypropyl)methacrylamide), synthetic oligonucleotides containing CpG or CpA motifs, monophosphoryl lipid A, MPL, Bacillus Clamette-Guerin, γ-interferon, Tissue Plasminogen Activator, LPS, Interleukin-1, Interleukin-2, UV light, a lymphotoxin, cachectin, a TNFR-2 agonist, an intracellular mediator of the TNF-alpha signaling

pathway, a NFκB inducing substance, IRF-1, STAT1, a lymphokine, a tumor necrosis factor-alpha (TNF-α) receptor II agonist, or the combination of TNF-alpha and an anti-TNFR-1 antibody.

22. (Withdrawn) The method of claim 21, wherein said TNF-alpha agonist or TNF-alpha inducing substance is Complete Freund's Adjuvant, Bacillus Clamette-Guerin, or γ-interferon.

23. (Withdrawn) The method of claim 14, wherein said organ or tissue is stimulated 6-12 hours before administering said composition.

24. (Withdrawn) A pluripotent cell transfected with a Hox11 gene, wherein said cell is capable of differentiating into a cell selected from the group consisting of: a pancreatic cell, a spleen cell, a liver cell, a kidney cell, and a bone cell.

25. (Withdrawn) The cell of claim 24, wherein said cell is capable of differentiating into a pancreatic cell.

26. (Withdrawn) The cell of claim 24, wherein said cell is transfected with a human Hox11 gene.

27. (Withdrawn) The cell of claim 24, wherein said pluripotent cell is derived from the spleen.

28. (Withdrawn) The cell of claim 24, wherein said pluripotent cell is derived from cord blood.

29. (Withdrawn) The cell of claim 24, wherein said pluripotent cell does not express CD45.

30. (Previously Presented) The method of claim 1, wherein said composition comprises cells that present MHC class I and peptide, wherein said MHC class I has at least one allele that matches an MHC class I allele expressed by said mammal.

31. (Withdrawn) A method for increasing or maintaining the number of functional cells of a predetermined type in an organ or tissue of a mammal, wherein said organ or tissue is injured, damaged, or deficient in said functional cells, said method comprising administering to said mammal an agent that induces and/or stimulates Hox11-expressing pluripotent cells.

32. (Withdrawn) The method of claim 31, wherein said Hox11-expressing cells are not bone marrow cells.

33. (Withdrawn) The method of claim 31, wherein said agent is a gene therapy vector comprising a Hox 11 gene operably linked to a promoter, wherein said vector expresses Hox 11 in said pluripotent cells.

34. (Withdrawn) The method of claim 31, said method further comprising: a) quantitating the number of said Hox11-expressing cells before administering said agent; and b) quantitating the number of said Hox11-expressing cells after administering said agent.

35. (Withdrawn) The method of claim 34, said method further comprising administering an additional amount of said agent if said number of Hox11-expressing cells of step a) is less than said number of step b).

36. (Withdrawn) The method of claim 34, said method further comprising administering a second agent that induces and/or stimulates pluripotent cells in which the Hox11 gene is expressed if said number of Hox11-expressing cells of step a) is less than said number of step b).

37. (Withdrawn) The method of claim 34, said method comprising: a) detecting in said mammal a first marker expressed by said Hox11-expressing cells and a second marker expressed by a control cell population selected from a non-pluripotent cell population or a second pluripotent cell population that is different from said Hox 11-expressing cells; b) quantitating the number of said Hox11-expressing cells and said control cells using said first marker and said second marker, respectively, before administering said composition; c) quantitating the number of said Hox11-expressing cells and said control cells using said first marker and said second marker, respectively, after administering said composition; and d) comparing the ratio of said Hox11-expressing cells to said control cells of step b) with the ratio of said Hox11-expressing cells to said control cells of step c).

38. (Withdrawn) The method of claim 37, said method further comprising administering an additional amount of said agent if said ratio of Hox11-expressing cells to control cells of step b) is less than said ratio of Hox11-expressing cells to control cells of step c).

39. (Withdrawn) The method of claim 37, said method further comprising administering a second agent that induces and/or stimulates pluripotent cells in which the Hox11 gene is expressed if said ratio of Hox11-expressing cells to control cells of step b) is less than said ratio of Hox11-expressing cells to control cells of step c).

40. (Withdrawn) The method of claim 37, wherein said first marker is the result of Hox11 gene expression.

41. (Withdrawn) The method of claim 37, wherein said first marker is detected by a compound that binds to said first marker with a binding constant (K_D) of less than or equal to 1 micromolar.

42. (Withdrawn) The method of claim 41, wherein said first marker is detected by an antibody.

43. (Withdrawn) The method of claim 31, wherein said agent is, or induces in said mammal, a cytokine, chemokine, or growth factor.

44. (Withdrawn) The method of claim 43, wherein said cytokine, chemokine, or growth factor is selected from the group consisting of: epidermal growth factor (EGF), platelet-derived growth factor (PDGF), fibroblast growth factors (FGFs), transforming growth factor-beta (TGF-β), transforming growth factor-alpha (TGF-α), vascular endothelial growth factor (VEGF), erythropoietin (Epo), insulin-like growth factor-I (IGF-I), insulin-like growth factor-II (IGF-II), interleukins, tumor necrosis factor-alpha (TNF-α), tumor necrosis factor-beta (TNF-β), interferon-gamma (INF-γ), stromal cell-derived factor-1 (SDF-1), and a colony stimulating factors (CSF).

45. (Previously Presented) The method of claim 1, wherein said organ or tissue is, or is part of, the pancreas, the spleen, the liver, the kidney, or the bone.

46. (Original) The method of claim 45, wherein said organ or tissue is, or is part of, the pancreas.

47. (Withdrawn) A method for increasing or maintaining the number of functional cells of a predetermined type in an organ or tissue of a mammal, wherein said organ or tissue is injured, damaged, or deficient in said functional cells, said method comprising administering to said mammal an agent that selectively inhibits, removes, or kills cell populations that interfere with or prevent the trafficking of, differentiation of, or growth of pluripotent cells.

48. (Withdrawn) The method of claim 47, wherein said agent targets a cell population deficient in the expression of CD180.

49. (Withdrawn) The method of claim 48, wherein said agent is BCG, LPS, a

triacetylated lipopeptide, phenol-soluble modulin, or OspA LP from B. burgdorferi, a triacetylated lipopeptide with TLR1 or TLR6, HSP60 with TL4, HSP60, a mannuronic acid polymer, a flavolipin, a tecihuronic acid, neumolysin, fimbriae, surfactant protein A, hyaluronan, heparin sulfate or a heparin sulfate fragment, a fibrinogen peptide, beta-defensin-2, flagellin, or imidazolquinoline.

50. (Withdrawn) The method of claim 47, wherein said pluripotent cells express Hox 11.

51. (Withdrawn) The method of claim 47, wherein said pluripotent cells are isogeneic.

52. (Withdrawn) The method of claim 47, wherein said pluripotent cells are semi-allogeneic.

53. (Previously Presented) The method of claim 1, further comprising administering to said mammal an agent that selectively inhibits, removes, or kills cell populations that interfere or prevent the trafficking of, differentiation of, or growth of Hox-11-expressing pluripotent cells.

54. (Currently Amended) The method of claim 53, wherein said cell populations comprise ~~blood cells are~~ lymphocytes.

55. (Original) The method of claim 53, wherein said agent comprises TNF-alpha.

56. (Original) The method of claim 53, wherein said agent comprises a TNF-alpha agonist or a TNF-alpha inducing substance.

57. (Previously Presented) The method of claim 56, wherein said TNF-alpha agonist or TNF-alpha inducing substance is Complete Freund's Adjuvant (CFA), ISS-ODN, microbial cell wall components with LPS-like activity, cholera particles, *E. coli* heat labile enterotoxin, *E. coli*

heat labile enterotoxin complexed with lecithin vesicles, ISCOMS-immune stimulating complexes, polyethylene glycol, poly(N-2-(hydroxypropyl)methacrylamide), synthetic oligonucleotides containing CpG or CpA motifs, monophosphoryl lipid A, Bacillus Clamette-Guerin, γ-interferon, Tissue Plasminogen Activator, LPS, Interleukin-1, Interleukin-2, UV light, a lymphotoxin, cachectin, a TNFR-2 agonist, an intracellular mediator of the TNF-alpha signaling pathway, a NFκB inducing substance, IRF-1, STAT1, a lymphokine, a tumor necrosis factor-alpha (TNF-α) receptor II agonist, or the combination of TNF-alpha and an anti-TNFR-1 antibody.

58. (Original) The method of claim 57, wherein said TNF-alpha agonist or TNF-alpha inducing substance is Complete Freund's Adjuvant, Bacillus Clamette-Guerin, or γ-interferon.

59. (Previously Presented) The method of claim 1, wherein said mammal has an autoimmune disease.

60. (Original) The method of claim 59, wherein said disease is diabetes.

61. (Original) The method of claim 59, wherein said disease is immunologically-mediated glomerulonephritis.

62. (Original) The method of claim 59, wherein said disease is chronic hepatitis, primary biliary cirrhosis, or primary sclerosing cholangitis.